

PLASMID-MEDIATED GENTAMICIN
RESISTANCE NOT ASSOCIATED
WITH KANAMYCIN RESISTANCE
IN ENTEROBACTERIACEAE

Sir :

R-Factors mediating resistance to aminoglycoside antibiotics have been shown to govern the synthesis of various drug-inactivating enzymes : a kanamycin-acetyl-transferase, a kanamycin-phospho-transferase, a streptomycin-adenylylate-synthetase, a streptomycin-phospho-synthetase and a gentamicin-adenylylate-synthetase^{1,14}). Recently, we obtained, from several hospitals, strains of *Enterobacter*, *Serratia*, *Klebsiella* and *Escherichia* sp. which were able to transfer into *E. coli* K12 a gentamicin resistance character different from the Gk character previously described in strain LA 290 R55^{15,16}).

Naturally occurring gentamicin resistant strains were conjugated to mutants of *E. coli* K12 resistant to sodium azothhydrate (LA 290) or to rifampicin (LA 534). Original strains and their R-factors are described in Table 1. Sensitivity to aminosidic antibiotics of *E. coli* recipients was determined by the agar dilution method¹⁷) using a STEER's apparatus (10⁴~10⁵ bacteria per spot). The MICs of gentamicin C, kanamycin A, tobramycin, paromomycin and lividomycin for *E. coli* J5 carrying R-factors listed in Table 1, are shown in Table 2. These strains were compared to reference strains : *E. coli* R5 kindly supplied by Dr. H. UMEZAWA which acetylates kanamycin and adenylates streptomycin ; *E. coli* R11-2 which phosphorylates kanamycin ; *E. coli* R55 which adenylates gentamicin (J. DAVIES, personal

Table 2. MIC in $\mu\text{g/ml}$ of deoxystreptamine antibiotics for *E. coli* J5 R⁺

R factor	Gen*	Tob.	Kan.	Par.	Liv.
R 135	32	0.25	1	2	4
R 136	64	0.25	1	4	4
R 137	32	0.25	1	2	4
R 138	32	0.25	1	2	4
R 140	64	0.25	1	2	4
R 141	128	0.25	1	4	8
R 146	32	0.25	1	4	4
Control					
R 55	64	32	64	4	4
R 11-2	0.25	0.25	512	>512	>512
R 5	0.5	8	16	4	8

Gen : gentamicin C ; Unilabo 3804, Tob : tobramycine ; Lilly XU 3740. Kan : kanamycin A ; Bristol 595. Paromomycin ; Parke Davis 10.254. Liv : lividomycin ; Roger Bellon.

* Levels for each fraction of gentamicin C : C1 Cla C2 kindly supplied by Dr. M. WEINSTEIN are similar.

communication).

E. coli harbouring R-factors R135, R136, R137, R138, R140, R141, R146 are resistant to gentamicin (32~128 $\mu\text{g/ml}$) but remain sensitive to kanamycin (1 $\mu\text{g/ml}$), tobramycin (0.25 $\mu\text{g/ml}$), paromomycin (2~4 $\mu\text{g/ml}$) and lividomycin (4~8 $\mu\text{g/ml}$). This type of resistance to deoxystreptamine antibiotics is clearly distinct from the one previously described in *E. coli* R55 inactivating gentamicin C, kanamycin A and tobramycin. Levels of resistance for streptomycin and spectinomycin when present are similar to those observed with strains adenylating streptomycin as *E. coli* R5.

R-Factor R135 (the first isolated), was transduced into *E. coli* K12 by phage P1 Kc ; the procedure has been described in a previous article¹⁵). Resistance determinants for tetracycline, sulfonamides, streptomycin and gentamicin were transduced simultaneously and are likely to belong to one single plasmid. Precise determination of inhibitory concentrations for deoxystreptamine antibiotics was done for *E. coli* transductant. Inhibitory concentrations (IC) 50 % values were calculated by a probit procedure¹⁸). Comparative results for sensitive strain *E. coli* J5, *E. coli* J5 R55 and

Table 1. Origin of R factors studies

Wild type strain	Origin	R factor	Transferred resistance characters*
<i>Enterobacter</i> 705	Foch hosp. Suresnes	R 135	T Su G S
<i>Klebsiella</i> 452	Foch hosp. Suresnes	R 136	A C T Su G
<i>Klebsiella</i> 12	C.H.U. Caen	R 138	A C T Su G S
<i>Serratia</i> 006	C.H.U. Angers	R 140	A C T Su G
<i>E. coli</i> 2423-1	C.H.U. Angers	R 141	A C T Su G S
<i>Klebsiella</i> 535	Claude Bernard hosp. Paris	R 137	A C T Su G S
<i>Enterobacter</i> 536	Claude Bernard hosp. Paris	R 146	A C T Su G S

* Resistance characters (disc method) : T : tetracycline ; Su : sulfonamides ; A : ampicillin ; C : chloramphenicol ; G : gentamicin ; S : streptomycin.

E. coli J5 R135 are summarized in Fig. 1. Differences in drug resistance specificity mediated by R55 adenylating gentamicin, and R135 are significant. There is actually a good correlation between drug resistance (MICs) and the inactivation of aminosidic drugs by bacterial enzymes¹⁹⁾ although it should be noted that N-acetylation due to R-factor R5 of gentamicin Cla, for instance, does not lead to real inactivation²⁰⁾.

The R-factor studied here mediates resistance to gentamicin C but not to kanamycin A and tobramycin (G character). These results suggest that the enzyme involved, if any, could be different from the one previously described and responsible for transferable gentamicin-kanamycin (Gk) resistance character.

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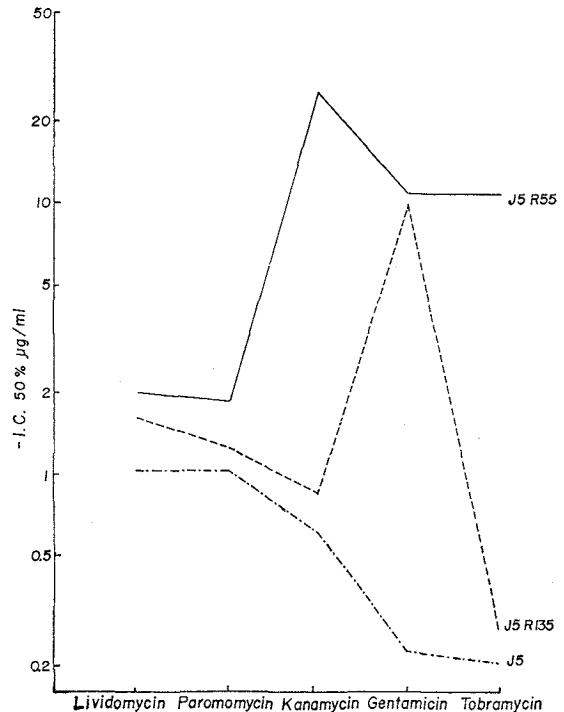
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Fig. 1. Inhibitory concentrations 50% in $\mu\text{g/ml}$ for *E. coli* J5 and *E. coli* J5 carrying R factors R135 and R55.



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